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# An optimized procedure for preconcentration, determination and on-line recovery of palladium using highly selective diphenyldiketone-monothiosemicarbazone modified silica gel

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# 1. Introduction

Palladium and its alloys have an extensive range of applications in various fields such as dentistry, metallurgy, instrument manufacturing, and chemical industries. The most significant and prevalent application of palladium is in the field of catalysis [1] and automotive catalytic converters [2] which leads to its accumulation in the environment. At the same time, palladium compounds are considered to be highly toxic and carcinogenic, and available evidences indicate that palladium is easily transported to biological material through plant roots, and ultimately intensified along the food chain [3]. The excessive exposure to palladium causes adverse health effects in humans such as primary skin problems, eye irritations, substantial degradation of DNA and cell mitochondria, and aggravation of hydroxyl radical damage and inhibition of the enzymes activity [4]. As the number of ecological and health problems associated with palladium contamination continues to rise, its determination and extraction is becoming very important in order to minimize its hazardous impact on human health and environment. But, in environmental matrices palladium is present at trace levels; therefore, there is a need to develop new sensitive analytical methods incorporating on-line separation and preconcentration steps, which not only make it feasible to

# ABSTRACT

selective. efficient novel highly and reusable chelating resin. diphenyldiketone-Α monothiosemicarbazone modified silica gel, was prepared and applied for the on-line separation and preconcentration of Pd(II) ions in catalytic converter and spiked tap water samples. Several parameters like effect of pH, sample volume, flow rate, type of eluent, and influence of various ionic interferences, etc. were evaluated for effective adsorption of palladium at trace levels. The resin was found to be highly selective for Pd(II) ions in the pH range 4–5 with a very high sorption capacity of 0.73 mmol/g and preconcentration factor of 335. The present environment friendly procedure has also been applied for large-scale extraction by employing the use of newly designed reactor in which on-line separation and preconcentration of Pd can be carried out easily and efficiently in short duration of time. © 2012 Elsevier B.V. All rights reserved.

> detect the metal ions but also separate the analytes from matrix interferences. Several analytical techniques such as coprecipitation, cloud point extraction, solvent extraction, electrodeposition and membrane filtration [5-13] have been employed for separation and preconcentration of Pd(II) ions. But, these techniques suffered from various drawbacks such as lack of sensitivity and selectivity, high cost, and use of large amount of toxic organic solvents which are harmful to human health and environment. Thus, solid phase extraction (SPE) [14-17] has emerged as an excellent separation technique in recent years in comparison to traditional extraction methodologies because of its numerous advantages of high enrichment factor, better separation, high selectivity and efficiency, ease of recovery and reusability, low cost because of lower consumption of reagents, less utilization of organic solvents and more importantly environment friendly. Furthermore, SPE also has the ability of easier integration into an automated analytical technique in on-line mode which has received significant interest during recent years, and is widely applied for selectivity and sensitivity enhancement. Thus, chelating resins, which can be easily prepared by immobilizing a metal specific chelating agent on the support material, are frequently used for preconcentration and separation of metal ions from interfering constituents. Among various support materials, silica gel is an extensively used support for various solid phase extractors because of its numerous advantages like high chemical and thermal stability, ready availability, economic viability, and easy and robust immobilization on its surface [18].

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Scheme 1. Preparation of DKTS-APSG resin.

In our ongoing contribution towards development of environment friendly strategy for extraction of metal ions using chelating resins [19–23], herein, we report a novel method for the selective recovery and determination of Pd(II) ions using silica based diphenyldiketone-monothiosemicarbazone chelating resin. Moreover, we have scaled up the procedure for on-line extraction by employing the use of newly designed reactor [24] in which on-line separation and preconcentration of Pd(II) ions can be carried out easily and effectively on large scale in short duration of time.

# 2. Experimental

#### 2.1. Instrumentation

FT-IR spectra were recorded on a Perkin–Elmer spectrometer at room temperature using KBr pellet technique in the range of 4000–400 cm<sup>-1</sup> under the atmospheric conditions with a resolution of 1 cm<sup>-1</sup>. The pH measurements were carried out using ELICO LI 120 pH meter. LABINDIA AA 7000 Atomic absorption spectrometer was employed for the determination of metal ion. The optimum

# and air flow rate, $1 \text{Lmin}^{-1}$ . The SEM images were recorded on ZEISS EVO 40 instrument. The surface area (BET) was measured with Gemini-V2.00 instrument (Micromeritics Instrument Corp.). Thermo gravimetric analysis was performed on DTG-60 Shimadzu instrument equipped with TG units at a heating rate of $10 \,^{\circ}\text{C/min}$ from $37 \,^{\circ}\text{C}$ to $600 \,^{\circ}\text{C}$ in N<sub>2</sub> atmosphere. Quantitative elemental analysis (CHN) was performed using Elementar Analysenysteme GmbhVarioEL.

parameters for Pd measurements are: wavelength, 244.4 nm; lamp

current, 3 mA; slit width, 0.2 nm; fuel flow rate, 1.5 Lmin<sup>-1</sup>,

### 2.2. Reagents and solutions

All the reagents used were of analytical grade. Silica gel and 3aminopropyltriethoxy silane (APTES) were procured from Sigma Aldrich (India). Diphenyldiketone and thiosemicarbazide were obtained from Spectrochem Pvt. Ltd. (India). Stock solutions of different metal ions containing 1000 mg/L of metal were used. All working solutions were prepared by stepwise dilution of stock



Scheme 2. Schematic diagram for on-line extraction and preconcentration of metal ions.

#### Table 1

Physico-chemical parameters of silica gel (SG), aminopropyl silica gel (APSG) and DKTS-APSG resin.

| Material   | Elemental analysis |      |      | Thermal analysis |                   | BET surface area (m²/g)  |        |
|------------|--------------------|------|------|------------------|-------------------|--------------------------|--------|
|            | %C                 | %Н   | %N   | %S               | $\Delta m/\%^{a}$ | $\Delta T/^{\circ}C^{b}$ |        |
| Silica gel | -                  | -    | -    | -                | -                 | -                        | 235.67 |
| APSG       | 6.60               | 2.38 | 2.20 | -                | 5.50              | 0-180                    | 152.04 |
|            |                    |      |      |                  | 2.80              | 180-600                  |        |
| DKTS-APSG  | 14.12              | 3.39 | 3.89 | 2.36             | 3.80              | 0-180                    | 83.66  |
|            |                    |      |      |                  | 19.00             | 180-600                  |        |

<sup>a</sup> Change in mass of material.

<sup>b</sup> Change in temperature.



**Fig. 1.** Electrostatic potential map of the monomeric unit (a perspective view) of DKTS–APSG.

solutions with double-distilled water, and their pH were adjusted using the buffer solutions.

# 2.3. Preparation of diphenyldiketone-monothiosemicarbazone (DKTS)

Diphenyldiketone-monothiosemicarbazone (DKTS) was prepared according to the reported procedure [25]. Equimolar amount of thiosemicarbazide (0.05 mol) and diphenyl diketone (0.05 mol) were dissolved in ethanol (250 ml) and refluxed for 4 h in presence of 2 ml of acetic acid. After the completion of reaction, orange precipitate of the product was filtered and recrystallized from ethanol.

# 2.4. Preparation of diphenylketone-monothiosemicarbazone modified silica (DKTS-APSG)

Aminopropyltriethoxysilane (1 ml) was dissolved in 100 ml of distilled water and acidified with acetic acid (pH 4). Then, 2 g of activated silica was added to the solution and stirred for 2 h at room temperature [26]. The product was filtered off and kept in oven

at 150 °C for 4 h. The obtained aminopropyl silica gel (APSG) was washed repeatedly with water, ethanol, and acetone for removal of un-reacted material, and then dried in the oven at 120 °C for 24 h. For immobilization of DKTS on APSG, 5 g of APSG was suspended in 50 ml of ethanol; 2 g of DKTS was then added to suspension and refluxed for 2 h. The orange colored crystalline solid obtained (DKTS–APSG) was filtered, washed with ethanol and dried under vacuum at 110 °C for 4 h (Scheme 1).

# 2.5. Sample preparation

The crushed and ground portion of the catalytic converter sample (0.25 g) was taken in beaker (100 ml) containing 25 ml of aqua-regia and the mixture was evaporated to nearly dryness. The white insoluble part was collected and dissolved in 100 ml water. Finally, the pH of the solution was adjusted to 4 with acetate buffer. For the analysis of tap water samples, water was taken directly from tap and spiked with Pd(II) ions. The resultant water sample was adjusted to required pH using buffer solution.

# 2.6. Analytical procedures

#### 2.6.1. Column method

A glass column (15 cm long), having a stopcock and a fritted porous disc (2.5 cm in diameter) was loaded with 50 mg of APSG–DKTS resin. It was washed with double-distilled water and conditioned with the buffer solution (sodium acetate/acetic acid) to the desired pH of 4. After conditioning, sample solution (10 ml) was passed in the column on to the resin bead at a flow rate of 10 ml/min. The adsorbed metal ion was then desorbed using 5 ml of eluting agent (mixture of HCl and thiourea) at a flow rate of 15 ml/min, and subjected to flame atomic absorption spectrometry for metal analysis using the optimum parameters. After every run, the resin in the column was washed with double distilled water and stored for the subsequent run.

## 2.6.2. Batch method

50 mg of APSG–DKTS resin was added to 250 ml glass stoppered bottle containing 10 ml of metal ion solution (5  $\mu$ g/ml). The solu-



**Fig. 2.** SEM images of fresh DKTS–APSG chelating resin, (a) at high magnification (26.11K×)(b) at low magnification (561×), and (c) the resin reused for 10 cycles (Magnification 408×).



**Fig. 3.** Effect of pH on adsorption of Al(III), Fe(II), Fe(III), Co(II), Zn(II), Cu(II) and Pd(II) onto DKTS-APSG resin. 50 mg resin, 10 ml test solution.

tion was adjusted to the desired pH of 4 and was kept on shaker for 30 min. The resin was then filtered, and concentration of the metal ion in the filtrate was determined by flame atomic absorption spectrophotometer using the same optimum parameters.

#### 2.6.3. Determination of adsorption capacity

For determination of adsorption capacity, 0.1 g of the resin was shaken with 50 ml of solution containing different concentrations of metal ion  $(10-500 \mu g/ml)$  in batch mode for 2 h at room temp. The filtrate from each flask was then subjected to AAS for the determination of metal ion present after requisite dilution. Adsorption capacity was calculated using the following equation  $Q = (C_o - C_e) V/w$ , where Q represents adsorption capacity (mg/g),  $C_o$  and  $C_e$  are the initial and final concentrations of metal ion (mg/l), w is the weight of the resin and V is the volume of metal ion solution.

#### 2.6.4. On-line separation of palladium

The on-line extraction and preconcentration system (Scheme 2) has been designed, which consists of the following units: (i) Batch extractor with a volume capacity of 5–10L fitted with an agitator, (ii) pH adjusting unit consisting of a pH controller with a probe electrode and buffer solution chambers, and (iii) Elution unit containing suction pump attached to the extractor through polypropylene tube in order to pump out the solution from extractor at a controlled flow rate. Initially, the batch extractor was charged with an optimized amount of silica supported chelating resin (DKTS–APSG), and then the metal ion solution was adjusted to required value with the help of pH adjusting unit, and thereafter the mixture in the extractor was subjected to agitation for an appropriate time. After the

#### Table 2

Influence of eluting agents on % recovery of the metal ions.

| Eluting agent                        | Volume for maximum<br>recovery (ml) | Recovery (%) |
|--------------------------------------|-------------------------------------|--------------|
| 0.1 M HCl                            | 5                                   | 40           |
| 0.2 M HCl                            | 5                                   | 45           |
| 1.0 M HCl                            | 6                                   | 50           |
| 2.0 M HCl                            | 5                                   | 60           |
| 0.5 M HNO3                           | 6                                   | 42           |
| 1.0 M HNO3                           | 6                                   | 50           |
| 1.0 M H <sub>2</sub> SO <sub>4</sub> | 6                                   | 44           |
| 0.1 M thiourea                       | 5                                   | 65           |
| 0.5 M thiourea                       | 6                                   | 75           |
| 1 M HCl+0.1 M thiourea               | 5                                   | 92           |
| 2 M HCl + 0.1 M thiourea             | 5                                   | 96           |
| 2 M HCl + 0.5 M thiourea             | 5                                   | 98.5         |

Table 3

Tolerance limit of the foreign ion species in presence of Pd(II) ions (5  $\mu$ g/ml).

| Foreign species                 | Tolerance limit (g/l) | Recovery (%) |
|---------------------------------|-----------------------|--------------|
| NaCl                            | 2.92                  | 94.4         |
| Na <sub>2</sub> SO <sub>4</sub> | 7.10                  | 93.3         |
| NaNO <sub>3</sub>               | 4.20                  | 95.0         |
| Na <sub>3</sub> PO <sub>4</sub> | 4.14                  | 94.0         |
| KCl                             | 3.72                  | 93.5         |
| KI                              | 1.60                  | 98.0         |
| CaCl <sub>2</sub>               | 1.10                  | 96.4         |
| MgCl <sub>2</sub>               | 4.76                  | 96.7         |
| NH <sub>4</sub> Cl              | 2.67                  | 95.5         |
| Cu <sup>2+</sup>                | 0.30                  | 96.4         |
| Cd <sup>2+</sup>                | 0.20                  | 92.8         |
| Zn <sup>2+</sup>                | 0.50                  | 95.0         |
| Cr <sup>3+</sup>                | 0.70                  | 98.0         |
| Co <sup>2+</sup>                | 0.70                  | 97.8         |
| Al <sup>3+</sup>                | 1.00                  | 98.3         |
| Fe <sup>2+</sup>                | 0.25                  | 96.3         |
| Mo <sup>6+</sup>                | 1.00                  | 98.6         |

adsorption of metal ion on the resin, the solution was pumped out through a polypropylene tube at the bottom of the extractor. Then for desorption of metal ion from the resin, selected eluting agent entered through the wash selection valve of the extractor for the elution. The concentration of metal ion was determined directly by aspirating the desorbed metal ion solution into atomic absorption spectrophotometer.

# 3. Result and discussion

## 3.1. Characterizations of APSG-DKTS resin

## 3.1.1. FT-IR spectroscopy

The grafting of APTES on silica gel and its subsequent functionalization with DKTS has been confirmed by FT-IR spectroscopy (Supplementary information). The peak at 990 cm<sup>-1</sup> in case of silica gel is assigned to silanol group. But in case of APSG, the peak due to silanol group has disappeared and the peak in the range of 2900–2800 cm<sup>-1</sup> due to  $-CH_2$  groups is observed which confirmed that APTES has been attached to silica gel. In the spectrum of grafted ligand the characteristic band of the imine group (C=N) appears at 1646 cm<sup>-1</sup>. The spectrum of the adsorbed resin after adsorbing Pd(II) ion shows a peak at 600 cm<sup>-1</sup> and also the frequency of C=N stretch frequency has been shifted compared to un-adsorbed resin confirming the covalent anchoring of DKTS on APSG.

#### 3.1.2. Elemental and thermal analysis

The quantitative estimation of modified surfaces (APSG and DKTS-APSG) was performed with elemental and thermogravimetric analyses (Table 1). The results of elemental analysis of APSG (nitrogen wt.%=2.20, carbon wt.%=6.60 hydrogen wt.%=2.38) gives rise to the grafting capacity of 1.57 mmol/g, and C/N ratio was found to be  $\sim$ 3 which is close to the expected value thereby confirming the 3-APTES grafting on silica gel. The loading of DKTS on APSG was found to be 0.74 mmol/g of silica using elemental results of APSG-DKTS (nitrogen wt.%=3.89, carbon wt.%=14.12 hydrogen wt.% = 3.39, sulfur wt.% = 2.36%). The presence of sulfur in the resin confirmed the grafting of DKTS on APSG. The TGA of the resin exhibited the mass loss of 3.8% in the first degradation assigned to absorbed water on the resin. In the second degradation, a mass loss of 19% was observed due to the decomposition of organic matter between 180 and 600 °C. The calculated loading of DKTS on APSG was found to be 0.74 mmol/g of silica which is in agreement with the results of elemental analysis.

#### Table 4

Comparison of important analytical characteristics of various support materials used for the separation and preconcentration of Pd(II) ions.

| Type of sorbent/ligand   | Support material           | Adsorption capacity<br>(mmol/g) | Preconcentration factor | LOD (ng/ml) | Ref.    |
|--|----------------------------|---------------------------------|-------------------------|-------------|---------|
| Dithiooxamide  | Polystyrene divinylbebzene | 0.10 mmol/g                     | _                       | -           | [27]    |
| Isodiphenylthiourea  | Silica gel                 | 0.18 mmol/g                     | _                       | 4.7         | [28]    |
| Thiophene-2-carbaldehyde   | Mesoporous silica          | 5 mg/g                          | 100                     | 0.20        | [29]    |
| N-allyl-N-propyl thiourea  | Silica gel                 | 0.8 mmol/g                      | 100                     | 0.05        | [30]    |
| Dimethyl glyoxime  | Silica gel                 | 4.06 mg/g                       | 75                      | 1200        | [31]    |
| Thioridazine HCl   | Octa-decyl silica          | -                               | 100                     | 12.0        | [32]    |
| Morin  | Silica gel                 | 0.24 mmol/g                     | 100                     | 210         | [33]    |
| Thiourea   | Silica gel                 | 0.29 mmol/g                     | 20                      | 21          | [34]    |
| Benzilmonopyridylhydrazone   | Silica gel                 | 0.65 mmol/g                     | 250                     | 0.1         | [35]    |
| BrPMAA (N-(4-bromophenyl)-2-methacryl amide)/AMPS  | _                          | -                               | 75                      | 1100        | [36]    |
| (acrylamido-2-methyl-1-propane<br>sulfonic acid)   |                            |                                 |                         |             |         |
| Thiourea   | Melamine formaldehyde      | 0.14 mmol/g                     | -                       | -           | [37]    |
| Polyethylineimine  | Alumina                    | 13 mg/g                         | 300                     | 0.042       | [38]    |
| Modified carbon nanotube   | -                          | 15.6 mg/g                       | 165                     | 0.30        | [39]    |
| p-Dimethylaminobenzylidene   | PEG (Poly ethylene glycol) | 0.09 mg/g                       | 125                     | 0.54        | [40]    |
| Iminodiacetic acid   | Silica gel                 | 0.19 mmol/g                     | 150                     | 800         | [41]    |
| Ion imprinted polymer  | -                          | 9.25 mg/g                       | -                       |             | [42]    |
| SFODME (solidified floating organic drop<br>micro extraction) based on USD (Ultra<br>sound dispersion) | -                          | -                               | 49.9                    | 0.6         | [43]    |
| 1-(2-Pyridylazo)-2-naphthol  | Nanoclay                   | 2.4 mg/g                        | 140                     | 0.1         | [44]    |
| [1,5-Bis(2-pyridyl)-3 sulphophenyl methylene thiocarbonohydrazide                                      | Dowex1X8-200               | _                               | 8.7                     | 2.0         | [45]    |
| Phenanthrenequinone-thiosemicarbazone  | Silica gel                 | 0.31 mmol/g                     | -                       | -           | [46]    |
| Diphenyldiketone-<br>monothiosemicarbazone   | Silica gel                 | 0.73 mmol/g                     | 335                     | 5.0         | Present |

# 3.1.3. BET surface area analysis

The anchoring of organic moieties onto the silica matrix obstructs the access of nitrogen gas molecules, thus reducing its surface area. As a result, as expected, the BET surface area decreased after grafting, according to the sequence SG > APSG > DKTS-APSG. The reduction in surface area in this order confirms the functionalization of silica gel with 3-aminopropyltriethoxy silane to give APSG, and its modification with DKTS to yield a chelating resin. The BET surface area of silica gel (SG), aminopropylsilica gel (APSG) and chelating resin DKTS-APSG were measured and presented in Table 1.

#### 3.1.4. Theoretical studies (molecular modeling)

In order to identify the binding sites on chelating resin (APSG–DKTS), molecular modeling was carried out using Spartan 08 software. The molecular geometry of the resin was optimized by Semi-Empirical (PM 3) method, and its electrostatic potential map (Fig. 1) was generated which provides insight into the nature of molecular recognition in ligand–metal bonding. The better coordinating sites for the metal are the ones which have relatively more negative electrostatic potential surface. Therefore, the available electron rich sites (red region) for binding of metal are N atom of C=N and S atom of C=S functionalities.

#### 3.1.5. Scanning electron microscopy (SEM)

The morphology of the chelating resin was characterized by SEM. SEM image at high magnification  $(26.11K \times)$  (Fig. 2a) clearly depicts the uniform dispersion of chelating agent on silica. During the preparation of chelating resin, the silica gel beads were subjected to continuous stirring but it is evident from SEM image at low magnification  $(561 \times)$  (Fig. 2b) that silica gel beads remain intact confirming its good mechanical strength. The SEM micrograph of reused (10 cycles) resin is also obtained (Fig. 2c) which confirms the stability of the prepared resin for its reusability even after several cycles.

### 3.2. Extraction studies

#### 3.2.1. Effect of pH

The effect of pH was found to be the most critical parameter for the adsorption of metal ions. The influence of pH on the adsorption of Al(III), Cr(III), Fe(III), Co(II), Fe(II), Zn(II), Pd(II), Cu(II) was investigated in the pH range of 2–9. Experimentation was carried out by passing 10 ml of test solution containing 5  $\mu$ g/ml of each analyte at different pH values. The percentage adsorption of different metal ions is shown in Fig. 3, and it was found that the resin showed maximum adsorption for Pd(II) ion in the pH range of 4–5. Hence, further studies were carried out for Pd(II) ion at pH 4.

#### 3.2.2. Selection of eluting agent

In order to investigate the most efficient eluting agent, first 10 ml of 5  $\mu$ g/ml Pd(II) ion solution was adsorbed on the resin and then eluted with varying concentration of HNO<sub>3</sub>, HCl, H<sub>2</sub>SO<sub>4</sub> and thiourea using different volumes (1–6 ml). Among these eluting agents, mixture of HCl and thiourea (5 ml) gave maximum recovery and hence selected as an appropriate eluting agent for further applications. The results are shown in Table 2.

#### 3.2.3. Reusability of the column

To ensure the stability and reusability of the resin, palladium was sorbed and desorbed on the resin in ten successive cycles. The column was reused after regeneration with 10 ml of mixture of thiourea (0.5 M) and HCl (2 M) and 100 ml distilled water, respectively, followed by vacuum drying. The adsorption capacity of the resin was found to be apparently constant even after its repeated use for 10 cycles. The surface morphology of the recycled resin was monitored with the help of scanning electron microscopy (SEM). The SEM micrograph of reused resin (10 cycles) shows the same morphology (Fig. 2c) as that of fresh resin (Fig. 2b) confirming its high mechanical stability. Thus, its high stability and reusability makes the present protocol environmentally benign.

#### 3.2.4. Effect of flow rate of metal ion solution and eluting agent

The flow rate of sample solution and eluting agent through the column is an imperative parameter for controlling the time of adsorption and analysis. The influence of flow rates were investigated in the range of 2–15 ml/min for Pd(II) ion solution, and it was observed that variable sample flow rate (rate at which sample solution is poured into the column) had no effect on the adsorption of metal ion up to 10 ml/min. At higher sample flow rates (>10 ml/min) the adsorption of metal ion was decreased by 20% (as increase in the flow rate allows less time for equilibration of metal ion with resin). Also, the eluent flow rate (rate at which the sample solution moves out of the column) had no effect up to 15 ml/min. Therefore, 10 ml/min was selected as sample flow rate and 15 ml/min was selected as flow rate for elution for further studies.

#### 3.2.5. Effect of sample volume and preconcentration factor

To enrich the low concentration of analytes from the large sample volume, preconcentration factor and effect of sample volume were determined. The effect of sample volume was studied by passing 250–2500 ml of metal solution through the column (fed with 0.1 g of resin) and keeping the total amount of loaded metal ion constant at 10 ng/ml and 20 ng/ml. The adsorbed metal ion was recovered by optimized eluting agent. The adsorption of metal ion was not affected by sample volume till 2000 ml. In the present work, the preconcentration factor was found to be 335 for 2000 ml of sample volume eluted with 6 ml of eluting agent.

## 3.2.6. Adsorption isotherm

To examine the adsorption behavior of resin, solution of metal ion in the range of 10–500 µg/ml were taken in the batches containing 0.1 g of chelating resin at room temperature. The concentration of un-adsorbed metal ion was determined by AAS. Langmuir equation,  $C_e/Q=1/Qb+C_e/Q_m$  was applied to determine the adsorption capacity of resin, where  $C_e$  is equilibrium concentration of metal ion, Q is the amount of metal ion adsorbed at equilibrium,  $Q_m$  is the maximum amount of adsorbed metal ion (76.92 mg/g), and *b* is the binding constant (0.26 L/mg). The amount of total metal sorbed on 1.0 g of resin is calculated from the plot obtained (Fig. 4).

#### 3.2.7. Effect of interfering ions

In order to examine the selectivity of the prepared resin for the determination and preconcentration of Pd(II) ions, effect of various electrolytes and metal ions were studied by varying the concentrations of added electrolytes from (1 to 10 g/L) in 10 ml solution of Pd(II) ions ( $5 \mu \text{g/ml}$ ). The effect of coexisting metal ions on the adsorption of Pd(II) ion was studied in binary mixtures as well as in multi-component mixture and no interference was observed up to the considerable concentration ( $100 \mu \text{g/ml}$ ). The tolerance limit (defined as the ion concentration causing a relative error of not more than 5%) of investigated electrolytes are given in Table 3, and found that many cations and anions, which are inevitably associated with Pd(II) ions up to an appreciable concentration.

# 3.2.8. Analytical performance (accuracy, precision and limit of detection)

The validity, accuracy and precision of the proposed method were tested by spiking known amount of metal ion, and it was found that the added and measured amount of analyte are in agreement with each other signifying the accuracy. The precision of the method under the optimum conditions was determined by performing 5 replica trials. The recoveries were found to be (>98%) with low relative standard deviation values (<2%). The obtained results (shown in Table 4) thus authenticate the accuracy and precision of the present procedure, and its interdependence from the matrix effect. For ten replicate measurements, LOD (Limit of detection, the



**Fig. 4.** (a) Adsorption isotherm of Pd(II) ions onto DKTS–APSG and (b) the linear form of the Langmuir adsorption isotherm. 100 mg resin, 50 ml test solution, pH 4.

amount of palladium required to give net signal equal to three times the standard deviation of signal for blank solution) was determined by passing 100 ml of solution through the column loaded with resin and fount to be 5 ng/ml. The equation of the calibration curve is Y = 0.0146X + 0.0061 with correlation coefficient of 0.9982, where Y is the absorbance and X is the concentration of the Pd(II) in  $\mu$ g mL<sup>-1</sup>.

#### 3.2.9. On-line separation

We have also performed the mentioned procedure for on-line separation and preconcentration of Pd(II) ions at bulk scale using newly designed reactor. On the basis of the aforementioned optimized conditions, 5 g of the resin was taken in the reactor and applied for online separation of palladium from the 2 L of metal ion solution (250  $\mu$ g/ml) with the help of the route elaborated in Section 2.6.4. The adsorption capacity was found in good agreement with the earlier determined value using column method on smaller scale. After the recovery of Pd(II) ions, the resin was washed, dried and recycled for next run.

Table 5

Analytical results for Pd (II) ions in tap water and catalytic converter samples.

| Concentration of palladium (µg) |  |  |  |  |
|---------------------------------|--|--|--|--|
| Added                           | Found (RSD)  | Recovery (%)   |  |  |
| 12.5                            | 12.10(1.25)  | 96.80  |  |  |
| 25.0                            | 24.65 (1.10)   | 98.60  |  |  |
| -                               | 33.12 (1.5) (without using resin)                      | -  |  |  |
| -                               | 30.11 (1.20) (using resin)                             | 98.05  |  |  |
| 25.0                            | 54.05 (1.15)   | 98.07  |  |  |
| 40.0                            | 69.50 (1.10)   | 99.10  |  |  |
|                                 | Concentr<br>Added<br>12.5<br>25.0<br>-<br>25.0<br>40.0 | Concentration of palladium (μg)   Added Found (RSD)   12.5 12.10 (1.25)   25.0 24.65 (1.10)   - 33.12 (1.5) (without using resin)   - 30.11 (1.20) (using resin)   25.0 54.05 (1.15)   40.0 69.50 (1.10) |  |  |

3.2.10. Comparison of the prepared resin with the literature precedents

Table 5 represents the literature precedents [27–46] of various sorbents for extraction of Pd(II) ions, and their analytical results have been compared with those obtained by using DKTS–APSG chelating resin. It is evident from the comparison of the results obtained that adsorption capacity and preconcentration factor of the present resin is very high making it superior to the reported ones in terms of selectivity, sensitivity and reusability [47].

## 4. Applications of the proposed method

# 4.1. Determination of Pd(II) ions in spiked tap water samples

For the preconcentration procedure, pH of water sample was adjusted to 4 and spiked with Pd(II) ions, and then subjected to the DKTS–APSG chelating resin. The sorbed Pd(II) ions were estimated using atomic absorption spectrometer. It was found that the resin was successful in quantitative extraction of the Pd(II) ions even in the presence of miscellaneous ions (Table 5).

#### 4.2. Determination of Pd(II) ions in catalytic converter sample

The applicability of the DKTS–APSG resin for the preconcentration of Pd(II) ions at trace level of was also tested using the catalytic converter sample. After the digestion of catalytic converter sample, the solution was adjusted to pH 4 and subjected to the DKTS–APSG chelating resin. The results are shown in Table 5.

# 5. Conclusion

We have developed an easy, efficient and environmental friendly methodology for the on-line determination of Pd(II) ions using a novel solid phase extractor, diphenyldiketonemonothiosemicarbazone functionalized silica gel. The proposed solid-phase extraction system for the first time enables an effective on-line palladium preconcentration and final determination of trace amount of Pd(II) ions in various samples with complex and variable matrices as a result of its excellent analytical characteristics such as low detection limit (5 ng/ml), high enrichment factor (335), high adsorption capacity (0.73 mmol/g) and good selectivity and precision. By using on-line solid phase extraction system, a large volume of samples (environmental/waste effluents) can be injected, and resin could continuously be used for a long period of time without any appreciable change in its sorption properties towards Pd(II) ions. Therefore, this is not only a practicable method for palladium trace analysis in a variety of matrices in order to reduce its hazardous impact on ecosystem but also a value addition because of the recovery of costly palladium metal.

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## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.jhazmat.2012.01.022.

#### References

- V. Polshettiwar, C. Len, A. Fihri, Silica-supported palladium: Sustainable catalysts for cross-coupling reactions, Coord. Chem. Rev. 253 (2009) 2599–2626.
- [2] R.J. Farrauto, R.M. Heck, Catalytic converters: state of the art and perspectives, Catal. Today 51 (1999) 351–360.
- [3] M.D. Shultz, J.P. Lassig, M.G. Gooch, B.R. Evans, J. Woodward, Palladium a new inhibitor of cellulase activities, Biochem. Biophys. Res. Commun. 209 (1995) 1046–1052.
- [4] J. Kielhorn, C. Melber, D. Keller, I. Mangelsdorf, Palladium- a review of exposure and effects to human health, Int. J. Hyg. Environ. Health 205 (2002) 417–432.
- [5] N. Ozturk, V.N. Bulut, C. Duran, M. Soylak, Coprecipitation of palladium(II) with 1,5-diphenylcarbazite-copper(II) and determination by flame atomic absorption spectrometry, Desalination 270 (2011) 130–134.
- [6] M. Soylak, M. Tuzen, Coprecipitation of gold(III), palladium(II) and lead(II) for their flame atomic absorption spectrometric determinations, J. Hazard. Mater. 152 (2008) 656–661.
- [7] M. Ghaedi, A. Shokrollahi, K. Niknam, E. Niknam, A. Najibi, M. Soylak, Cloud point extraction and flame atomic absorption spectrometric determination of cadmium(II), lead(II), palladium(II) and silver(I) in environmental samples, J. Hazard. Mater. 168 (2009) 1022–1027.
- [8] D.L.G. Borges, M.A.M.S.D. Veiga, V.L.A. Frescura, B. Welz, A.J. Curtius, Cloudpoint extraction for the determination of Cd, Pb and Pd in blood by electrothermal atomic absorption spectrometry, using Ir or Ru as permanent modifiers, J. Anal. At. Spectrom. 18 (2003) 501–507.
- [9] B.K. Priya, P. Subrahmanayam, K. Suvardhan, K.S. Kumar, D. Rekha, A.V. Rao, G.C. Rao, P. Chiranjeevi, Cloud point extraction of palladium in water samples and alloy mixtures using new synthesized reagent with flame atomic absorption spectrometry, J. Hazard. Mater. 144 (2007) 152–158.
- [10] J.Y. Lee, B. Raju, B.N. Kumar, J.R. Kumar, H.K. Park, B.R. Reddy, Solvent extraction separation and recovery of palladium and platinum from chloride leach liquors of spent automobile catalyst, Sep. Purif. Technol. 73 (2010) 213–218.
- [11] L. Pan, Z. Zhang, Solvent extraction and separation of palladium(II) and platinum(IV) from hydrochloric acid medium with dibutyl sulfoxide, Miner. Eng. 22 (2009) 1271–1276.
- [12] M. Vaezzadeh, F. Shemirani, B. Majidi, Microextraction technique based on ionic liquid for preconcentration and determination of palladium in food additive, sea water, tea and biological samples, Food Chem. Toxicol. 48 (2010) 1455–1460.
- [13] J. Komarek, P. Krasensky, J. Balcar, P. Rehulka, Determination of palladium and platinum by electrothermal atomic absorption spectroscopy after deposition on a graphite furnace, Spectrochim. Acta Part B 54 (1999) 739–743.
- [14] R.K. Sharma, S. Mittal, M. Koel, Analysis of trace amounts of metal ions using silica-based chelating resins: a green analytical method, Crit. Rev. Anal. Chem. 33 (2003) 183–197.
- [15] L. Elci, M. Soylak, E.B. Buyuksekerci, Separation of gold, palladium and platinum from metallurgical samples using an Amberlite XAD-7 resin column prior to their atomic absorption spectrometric determinations, Anal. Sci. 19 (2003) 1621–1624.
- [16] M. Soylak, L. Elci, A Sorbent extraction procedure for the preconcentration of gold, silver and palladium on an activated carbon column, Anal. Lett. 33 (2000) 513–525.
- [17] V.N. Bulut, M. Tufekci, C. Duran, M. Soylak, H. Kantekin, Selective solid phase extraction for separation and preconcentration of palladium from gold ore and anode slime after complexation with a N<sub>4</sub>O<sub>2</sub> mixed donor ligand derivative, clean-soil, air, water 38 (2010) 678–683.
- [18] R. Sharma, S. Dhingra, Designing and Synthesis of Functionalized Silica Gels and their Applications as Metal Scavengers, Sensors, and Catalysts: A Green Chemistry Approach, LAP Lambert Academic Publishing, Germany, 2011.
- [19] R.K. Sharma, Design, synthesis, and application of chelating polymers for separation and determination of trace and toxic metal ions: a green analytical method, Pure Appl. Chem. 73 (2001) 181–186.
- [20] R.K. Sharma, A. Goel, Development of a Cr(III)-specific potentiometric sensor using aurin tricarboxylic acid modified silica, Anal. Chim. Acta 534 (2005) 137–142.
- [21] R.K. Sharma, S. Mittal, S. Azmi, A. Adholeya, Surface modified silica gel for extraction of metal ions: an environment friendly method for waste treatment, Surf. Eng. 21 (2005) 232–237.

- [22] R.K. Sharma, P. Pant, Preconcentration and determination of trace metal ions from aqueous samples by newly developed gallic acid modified Amberlite XAD-16 chelating resin, J. Hazard. Mater. 163 (2009) 295–301.
- [23] R.K. Sharma, P. Pant, Solid phase extraction and determination of metal ions in aqueous samples using Quercetin modified Amberlite XAD-16 chelating polymer as metal extractant, Int. J. Environ. Anal. Chem. 89 (2009) 503–514.
- [24] A. Adholeya, R.K. Sharma, Patent: CBR 4171 1146/del/2010.
- [25] O.E. Offiong, Study of the stereochemistry of diphenyldiketone monothiosemicarbazone and its transition metal complex, Transit. Met. Chem. 20 (1995) 126–131.
- [26] A.M. Donia, A.A. Atia, W.A. Al-amrani, A.M. El-Nahas, Effect of structural properties of acid dyes on their adsorption behaviour from aqueous solutions by amine modified silica, J. Hazard. Mater. 161 (2009) 1544–1550.
- [27] S. Dutta, P.K. Mohapatra, V.K. Manchanda, A.K. Das, Preconcentration and separation of trace amounts of palladium using dithiooxamide functionalized chelating resin followed by its determination using radiotracer technique, J. Radioanal. Nucl. Chem. 274 (2007) 245–250.
- [28] P. Liu, Z. Su, X. Wu, Q. Pu, Application of isodiphenylthiourea immobilized silica gel to flow injection on-line microcolumn preconcentration and separation coupled with flame absorption spectrometry for interference-free determination of trace silver, gold, palladium and platinum in geological and metallurgical samples, J. Anal. At. Spectrom. 17 (2002) 125–130.
- [29] M.R. Jamali, Y. Assadi, F. Shemirani, M.S. Niasari, Application of thiophene-2-carbaldehyde-modified mesoporous silica as a new sorbent for separation and preconcentration of palladium prior to inductively coupled plasma atomic emission spectrometric determination, Talanta 71 (2007) 1524–1529.
- [30] V.N. Losev, G.V. Volkova, N.V. Maznyak, A.K. Trofimchuk, E.Y. Yanovskaya, Palladium adsorption on silica modified with *N*-allyl-*N*-propylthiourea and subsequent determination by spectrometry, J. Anal. Chem. 54 (1999) 1109–1113.
- [31] S. Tokalioglu, T. Oymak, S. Kartal, Determination of palladium in various samples by atomic absorption spectrometry after preconcentration with dimethylglyoxime on silica gel, Anal. Chim. Acta 511 (2004) 255–260.
- [32] K. Farhadi, G. Teimouri, Flame atomic absorption determination of palladium in solutions after preconcentration using octadecyl silica membrane disks modified by thioridazine HCl, Talanta 65 (2005) 925–929.
- [33] M.M. Hassanien, K.S.A. Sherbini, Synthesis and characterization of morinfunctionalised silica gel for the enrichment of some precious metal ions, Talanta 68 (2006) 1550–1559.
- [34] P. Liu, Q. Pu, Z. Su, Synthesis of silica gel immobilized thiourea and its application to the on-line preconcentration and separation of silver gold and palladium, Analyst 125 (2006) 147–150.

- [35] M.M. Hassanien, FAAS determination of palladium after its selective recovery by silica modified with hydrazone derivative, Microchim. Acta 167 (2009) 81–89.
- [36] S. Tokalıoglu, V. Yılmaz, S. Kartal, A. Delibas, C. Soykan, Solid phase extraction of Pd(II) on a newly synthesized chelating resin prior to determination by flame atomic absorption spectrometry, Microchim. Acta 165 (2009) 347–352.
- [37] E. Birinci, M. Gulfen, A. Aydin, Separation and recovery of palladium(II) from base metal ions by melamine-formaldehyde-thiourea (MFT) chelating resin, Hydrometall. 95 (2009) 15–21.
- [38] F. Sabermahani, M.A. Taher, Flame atomic absorption and determination of palladium after separation and preconcentration using polyethyleneimine water-soluble polymer/Alumina as a new sorbent, J. Anal. At. Spectrom. 25 (2010) 1102–1106.
- [39] C. Yuan, Y. Zhang, S. Wang, A. Chang, Separation and preconcentration of palladium using modified multi-walled carbon nanotube without chelating agent, Microchim. Acta 173 (2011) 361–367.
- [40] S. Rastegarzadeh, N. Pourreza, A.R. Kiasat, H. Yahyavi, Selective solid phase extraction of palladium by adsorption of its 5-(p-dimethylaminobenzylidene) rhodanine complex on silica-PEG as a new adsorbent, Microchim. Acta 170 (2010) 135–140.
- [41] G. Venkatesh, A.K. Singh Enrichment, flame atomic absorption spectrometric determination of palladium using chelating matrices designed by functionalizing Amberlite XAD-2/16 and silica gel, Microchim. Acta 159 (2007) 149–155.
- [42] B.G. Zylkiewicz, B. Lesniewska, I. Wawreniuk, Assessment of ion imprinted polymers based on Pd(II) chelate complexes for preconcentration and FAAS determination of palladium, Talanta 83 (2010) 596–604.
- [43] M. Mohamadi, A. Mostafavi, A novel solidified floating organic drop microextraction based on ultrasound-dispersion for separation preconcentration of palladium in aqueous samples, Talanta 81 (2010) 309–313.
- [44] D. Afzali, A. Mostafavi, Z. Afzali, Determination of trace amounts of Pd(II) ions in water and road dust samples by flame atomic absorption spectrometry after preconcentration on modified organo nanoclay, J. AOAC Int. 93 (2010) 1952–1956.
- [45] C.B. Ojeda, F.S. Rojas, J.M.C. Pavon, On-line preconcentration of palladium(II) using a microcolumn packed with a chelating resin, and its subsequent determination by graphite furnace atomic absorption spectrometry, Microchim. Acta 158 (2007) 103–110.
- [46] M.M. Hassanien, S. Khaled, Selective separation of palladium(II) from precious metal ions using thiosemicarbazone derivatives from acidic media by solid phase and solvent extractions. Desalination Water Treat. 16 (2010) 329–338.
- [47] H. Ebrahimzadeh, N. Tavassoli, M.M. Amini, Y. Fazaeli, H. Abedi, Determination of very low levels of gold and palladium in wastewater and soil samples by atomic absorption after preconcentration on modified MCM-48 and MCM-41 silica, Talanta 81 (2010) 1183–1188.